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<b>(21) International Application Number:</b> PCT/US00/07148 <b>(22) International Filing Date:</b> 17 March 2000 (17.03.00)  <b>(30) Priority Data:</b> 09/271,970      18 March 1999 (18.03.99)      US  <b>(71) Applicant (for all designated States except US):</b> XOMA TECHNOLOGY LTD. [—/—]; c/o XOMA (US) LLC, 2910 Seventh Street, Berkeley, CA 94710 (US).  <b>(72) Inventors; and</b> <b>(75) Inventors/Applicants (for US only):</b> BETTER, Marc, D. [US/US]; 2462 Zorada Drive, Los Angeles, CA 90046 (US). GAVIT, Patrick, D. [US/US]; 522 North Larkin Drive, Covina, CA 91722 (US).  <b>(74) Agent:</b> MCNICHOLAS, Janet, M.; McAndrews, Held & Malloy, Suite 3400, 500 West Madison, Chicago, IL 60661 (US).		<b>(81) Designated States:</b> AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, DM, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).  <b>Published</b> <i>With international search report.</i> <i>Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i>
<b>(54) Title:</b> IMPROVED METHODS FOR RECOMBINANT PEPTIDE PRODUCTION  <b>(57) Abstract</b> <p>The present invention provides improved methods for the production of recombinant peptides from bacterial cells.</p>		

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## IMPROVED METHODS FOR RECOMBINANT PEPTIDE PRODUCTION

The present invention relates generally to improved methods for the production of recombinant peptides from bacterial cells.

5

### BACKGROUND OF THE INVENTION

Although bioactive peptides can be produced chemically by a variety of synthesis strategies, recombinant production of peptides, including those in the 5-50 amino acid size range, offers the potential for large scale production at reasonable cost. However, expression of very short polypeptide chains can sometimes be problematic in microbial systems, including in bacterial cells such as *Escherichia coli*. This is true even when the peptide sequence is expressed as part of a fusion protein. As part of a fusion protein, peptides may be directed to specific cellular compartments, *i.e.* cytoplasm, periplasm, or media, with the goal of achieving high expression yield and avoiding cellular degradative processes.

Preparation of a peptide from a fusion protein in pure form requires that the peptide be released and recovered from the fusion protein by some mechanism and then obtained by isolation or purification. Methods for cleaving fusion proteins have been identified. Each method recognizes a chemical or enzymatic cleavage site that links the carrier protein to the desired protein or peptide [Forsberg et al., *I. J. Protein Chem.* 11, 201-211, (1992)]. Chemical cleavage reagents in general recognize single or paired amino acid residues which may occur at multiple sites along the primary sequence, and therefore may be of limited utility for release of large peptides or protein domains which contain multiple internal recognition sites. However, recognition sites for chemical cleavage can be useful at the junction of short peptides and carrier proteins. Chemical cleavage reagents include cyanogen bromide, which cleaves at methionine residues [Piers et al., *Gene*, 134, 7, (1993)], N-chloro succinimide [Forsberg et al., *Biofactors* 2, 105-112, (1989)] or BNPS-skatole [Knott et al., *Eur. J. Biochem.* 174, 405-410, (1988); Dykes et al., *Eur. J. Biochem.* 174, 411-416, (1988)] which cleaves at tryptophan residues, dilute acid which cleaves aspartyl-prolyl bonds [Gram et al., *Bio/Technology* 12, 1017-1023, (1994);

Marcus, *Int. J. Peptide Protein Res.*, 25, 542-546, (1985)], and hydroxylamine which cleaves asparagine-glycine bonds at pH 9.0 [Moks et al., *Bio/Technology* 5, 379-382, (1987)].

Of interest is U.S. Patent No. 5,851,802 which describes a series of  
5 recombinant peptide expression vectors that encode peptide sequences derived from bactericidal/permeability-increasing protein (BPI) linked via amino acid cleavage site sequences as fusions to carrier protein sequences. In some fusion protein constructs, an acid labile aspartyl-prolyl bond was positioned at the junction between the peptide and carrier protein sequences. BPI-derived peptides  
10 were released from the fusion proteins by dilute acid treatment of isolated inclusion bodies without prior solubilization of the inclusion bodies. The released peptides were soluble in the aqueous acidic environment. In addition, BPI-derived peptides were obtained from fusion proteins under conditions where the fusion proteins were secreted into the culture medium. Those secreted fusion  
15 proteins were then purified and treated with dilute acid to release the peptide.

Of additional interest are the disclosures of the following references which relate to recombinant fusion proteins and peptides.

Shen, *Proc. Nat'l. Acad. Sci. (USA)*, 281, 4627 (1984) describes bacterial expression as insoluble inclusion bodies of a fusion protein encoding pro-insulin and  
20  $\beta$ -galactosidase; the inclusion bodies were first isolated and then solubilized with formic acid prior to cleavage with cyanogen bromide.

Kempe et al., *Gene*, 39, 239 (1985) describes expression as insoluble inclusion bodies in *E. coli* of a fusion protein encoding multiple units of neuropeptide substance P and  $\beta$ -galactosidase; the inclusion bodies were first  
25 isolated and then solubilized with formic acid prior to cleavage with cyanogen bromide.

Lennick et al., *Gene*, 61, 103 (1987) describes expression as insoluble inclusion bodies in *E. coli* of a fusion protein encoding multiple units (8) of  $\alpha$ -human atrial natriuretic peptide; the inclusion bodies were first isolated and then solubilized  
30 with urea prior to endoproteinase cleavage.

Dykes et al., *Eur. J. Biochem.*, 174, 411 (1988) describes soluble intracellular expression in *E. coli* of a fusion protein encoding  $\alpha$ -human atrial natriuretic peptide

and chloramphenicol acetyltransferase; the fusion protein was proteolytically cleaved or chemically cleaved with 2-(2-nitrophenylsulphenyl)-E-methyl-3'-bromoindolenine to release peptide.

Ray *et al.*, *Bio/Technology*, 11, 64 (1993) describes soluble intracellular  
5 expression in *E. coli* of a fusion protein encoding salmon calcitonin and glutathione-S-transferase; the fusion protein was cleaved with cyanogen bromide.

Schellenberger *et al.*, *Int. J. Peptide Protein Res.*, 41, 326 (1993) describes expression as insoluble inclusion bodies of a fusion protein encoding a substance P peptide (11 a.a.) and  $\beta$ -galactosidase; the inclusion bodies were first isolated and then  
10 treated with chymotrypsin to cleave the fusion protein.

Hancock *et al.*, WO94/04688 (PCT/CA93/00342) and Piers *et al.* (Hancock), *Gene*, 134, 7 (1993) describe (a) expression as insoluble inclusion bodies in *E. coli* of a fusion protein encoding a defensin peptide designated human neutrophil peptide 1 (HNP-1) or a hybrid cecropin/mellitin (CEME) peptide and glutathione-5-  
15 transferase (GST); the inclusion bodies were first isolated and then: (i) extracted with 3% octyl-polyoxyethylene prior to urea solubilization and prior to factor X<sub>a</sub> protease for HNP1-GST fusion protein or (ii) solubilized with formic acid prior to cyanogen bromide cleavage for CEME-GST fusion protein; (b) expression in the extracellular supernatant of *S. aureus* of a fusion protein encoding CEME peptide  
20 and protein A; (c) proteolytic degradation of certain fusion proteins with some fusion protein purified; and (d) proteolytic degradation of other fusion proteins and inability to recover and purify the fusion protein.

Lai *et al.*, U.S. Patent No. 5,206,154 and Callaway, Lai *et al.* *Antimicrob. Agents & Chemo.*, 37:1614 (1993) describe expression as insoluble inclusion bodies  
25 of a fusion protein encoding a cecropin peptide and the protein encoded by the 5'-end of the L-ribulokinase gene; the inclusion bodies were first isolated and then solubilized with formic acid prior to cleavage with cyanogen bromide.

Gramm *et al.*, *Bio/Technology*, 12:1017 (1994) describes expression as insoluble inclusion bodies in *E. coli* of a fusion protein encoding a human  
30 parathyroid hormone peptide and a bacteriophage T4-encoded gp55 protein; the inclusion bodies were first isolated (6% wt/vol.) and then were treated with acid to hydrolyze the Asp-Pro cleavage site.

Kuliopulos *et al.*, *J. Am. Chem. Soc.*, 116:4599 (1994) describes expression as insoluble inclusion bodies in *E. coli* of a fusion protein encoding multiple units of a yeast  $\alpha$ -mating type peptide and a bacterial ketosteroid isomerase protein; the inclusion bodies were first isolated and then solubilized with guanidine prior to  
5 cyanogen bromide cleavage.

Pilon *et al.*, *Biotechnol. Prog.*, 13, 374-379 (1997) describe soluble intracellular expression in *E. coli* of a fusion protein encoding a peptide and ubiquitin; the fusion protein was cleaved with a ubiquitin specific protease, UCH-L3.

Haught *et al.*, *Biotechnol. Bioengineer.*, 57, 55-61 (1998) describe expression  
10 as insoluble inclusion bodies in *E. coli* of a fusion protein encoding an antimicrobial peptide designated P2 and bovine prochymosin; the inclusion bodies were first isolated and then solubilized with formic acid prior to cleavage with cyanogen bromide.

The above-references indicate that production of small peptides from bacteria  
15 has been problematic for a variety of reasons. Proteolysis of some peptides has been particularly problematic, even where the peptide is made as a part of a larger fusion protein. Such fusion proteins comprising a carrier protein/peptide may not be expressed by bacterial host cells or may be expressed but cleaved by bacterial proteases. In particular, difficulties in expressing cationic antimicrobial peptides in  
20 bacteria have been described by Hancock *et al.* WO94/04688 (PCT/CA93/00342) referenced above, due in their view to the susceptibility of such polycationic peptides to bacterial protease degradation.

The production of peptide for preclinical and clinical evaluation often requires multigram quantities [Kelley, *Bio/Technology* 14, 28-31 (1996)]. If  
25 production of recombinant peptides can be achieved at this large scale, such production can potentially be economical. However, downstream processing steps for the production of peptides and proteins from bacteria can often contribute a significant fraction of total production cost. Initial recovery of peptide from bacterial inclusion bodies of fusion proteins, for example, generally  
30 requires multiple distinct processing steps, including the following four steps: (1) cell disruption/lysis, (2) isolation of inclusion bodies from the disrupted/lysed cells, (3) solubilization of the isolated inclusion bodies in denaturant or detergent

to obtain solubilized fusion protein, and (4) fusion protein cleavage and separation of peptide and carrier protein. It is desirable that aspects of the recombinant production process be improved and/or optimized in order to make large-scale production of peptides by recombinant means more economically viable.

5           There continues to exist a need in the art for improved methods for recombinant production of peptides from bacterial cells, particularly for simpler methods that do not require a multiplicity of steps, including, for example, the step of isolation or purification of peptide fusion proteins or the step of isolation or purification of inclusion bodies comprising the fusion proteins in order to obtain  
10       the recombinant peptide.

### SUMMARY OF THE INVENTION

          The present invention provides improved methods for the production of recombinant peptides from bacterial cells. The improved methods preclude the  
15       need for the isolation and solubilization of inclusion bodies or the isolation and purification of peptide fusion proteins. The improved methods accomplish cell disruption/lysis and release of peptide from bacterial cells or bacterial cell cultures in a single step. Fusion proteins useful in methods of the invention comprise at least one peptide sequence, a carrier protein sequence, and at least one acid-  
20       sensitive amino acid cleavage site sequence located between the peptide sequence and the carrier protein sequence. The invention provides improved methods for the microbial production of peptides from such fusion proteins expressed intracellularly in bacterial cells. The recombinant peptides recovered according to the invention are released by acid cleavage at the acid-sensitive cleavage site(s) in  
25       the fusion protein. Recombinant peptides are thus efficiently and economically produced according to the invention.

          The invention thus provides an improved method for obtaining a peptide from bacterial cells after expression inside the cells of a fusion protein, wherein the fusion protein comprises the peptide, a carrier protein and an acid-cleavable  
30       site between the peptide and the carrier protein, with the improvement comprising treating the bacterial cells with acid under conditions sufficient in a single step to disrupt or lyse the cells and release the peptide from the fusion protein. An

improved method may include the additional step of obtaining the released peptide separated from the disrupted or lysed cells. According to the invention, the released peptide may be separated from the disrupted or lysed cells by a separation device, such as a centrifugation device or a filtration device. The invention also provides an improved method for obtaining a peptide from bacterial cells after expression inside the cells of a fusion protein, wherein the fusion protein comprises the peptide, a carrier protein and an acid-cleavable site between the peptide and the carrier protein with the improvement comprising the following steps: (a) treating the bacterial cells with acid under conditions sufficient to  
5 disrupt or lyse the cells and release the peptide from the fusion protein; (b) separating soluble material from insoluble material after step (a); and (c) recovering the released peptide in the soluble material after step (b). According to the invention, the soluble material may be separated from the insoluble material by a separation device, such as a centrifugation device or a filtration device.  
10 Improved methods of the invention may be employed where the bacterial cells are in cell culture media for the acid treatment, or where the bacterial cells have been separated from cell culture media for the acid treatment, or where the bacterial cells are in cell culture media in a fermentation vessel for the acid treatment. According to methods of the invention, preferred acid-cleavable sites in the fusion  
15 protein include an Asp-Pro cleavage site. Preferably, the carrier protein is expressed as an insoluble protein inside the bacterial cells.

Improved methods of the invention for recombinant microbial production of peptides from fusion proteins are based on the surprising discovery that bacterial cell disruption/lysis and peptide release may be accomplished simultaneously in a single  
25 step. According to the invention, no process step is required for the isolation from the cells of inclusion bodies and solubilization of such inclusion bodies prior to peptide release and recovery. Similarly, no process step is required for the purification of the fusion proteins expressed in large amounts intracellularly as soluble or insoluble proteins in bacterial host cells prior to peptide release and  
30 recovery. Remarkably, peptides efficiently produced as components of fusion proteins by the bacterial host cells are efficiently cleaved and released from the fusion proteins by a single step of cell disruption/lysis and peptide release. It is



particularly surprising that peptides according to the invention effectively made in *E. coli* are released in soluble form in this single step of cell disruption/lysis and peptide cleavage and are easily recovered from insoluble cell material. By way of example, recombinant BPI-derived peptides having one or more of the biological activities of BPI (*e.g.*, LPS binding, LPS neutralization, heparin binding, heparin neutralization, antimicrobial activity) have been produced and recovered according to the methods of invention. Thus, the invention provides improved methods of bacterial cell production of functional recombinant peptides.

10

### DETAILED DESCRIPTION

The present invention provides improved recombinant peptide production methods. Recombinant peptides encoded by and released from fusion proteins are recovered according to these improved methods. Fusion proteins useful in methods according to the invention comprise a peptide sequence, a carrier protein sequence and an acid-sensitive amino acid cleavage site sequence between the peptide and carrier protein sequences. Improved methods according to the invention accomplish cell disruption/lysis and release of peptide from the cells in a single step using bacterial cells or bacterial cell cultures (*e.g.*, fermentation cultures). The methods preclude the need for disruption/lysis followed by isolation and solubilization of inclusion bodies of the fusion proteins from the bacterial cells prior to peptide release and recovery. Unexpectedly, single step treatment of bacterial cells or bacterial cell cultures under conditions of acid pH and temperature sufficient to cleave and release peptides simultaneous with cell disruption/lysis, allows the direct recovery of soluble peptide from insoluble cell lysis material. Fusion proteins containing BPI-derived peptides with antimicrobial activity were expressed intracellularly in large amounts without significant proteolysis, until acidification of the bacterial cells. A variety of BPI-derived peptides, including those comprising the sequences listed in Table 4 of U.S. Patent No. 5,851,802 incorporated by reference herein in its entirety, may be produced by recombinant methods according to the invention.

An advantage provided by the present invention is the ability to produce peptides from fusion proteins more efficiently and economically from bacterial host

cells. Additional advantages include the ability to recover and obtain homogeneous peptide in large amounts via improved methods that are particularly amenable to scale-up in large fermentation vessels.

"BPI-derived peptide" or "BPI peptide" as used herein refers to a peptide  
5 derived from or based on bactericidal/permeability-increasing protein (BPI),  
including peptides derived from Domain I (amino acids 17-45), Domain II (amino  
acids 65-99) and Domain III (amino acids 142-169) of BPI (SEQ ID NOS: 15 and  
16), each peptide having an amino acid sequence that is the amino acid sequence of a  
BPI functional domain or a subsequence thereof and variants of the sequence or  
10 subsequence having at least one of the biological activities of BPI. The amino acid  
sequence of the entire human BPI protein and the nucleic acid sequence of DNA  
encoding the protein have been reported in Figure 1 of Gray et al., *J. Biol. Chem.*,  
264, 9505 (1989), incorporated herein by reference. The Gray et al. DNA and amino  
acid sequences are set out in SEQ ID NOS: 15 and 16 hereto. An N-terminal BPI  
15 fragment of approximately 23 kD, referred to as rBPI<sub>23</sub>, [Gazzano-Santoro et al.,  
*Infect. Immun.* 60, 4754-4761 (1992)], an analog designated rBPI<sub>21</sub> or rBPI<sub>21</sub>Δcys  
(U.S. Patent No. 5,420,019, incorporated by reference herein) as well as recombinant  
holoprotein, also referred to as rBPI, have been produced having sequences set out in  
SEQ ID NOS: 15 and 16, except that valine at position 151 is specified by GTG  
20 rather than GTC and residue 185 is glutamic acid (specified by GAG) rather than  
lysine (specified by AAG). As used herein, a "biological activity of BPI" refers to  
LPS binding, LPS neutralization, heparin binding, heparin neutralization or  
antimicrobial activity (including anti-bacterial and anti-fungal activity). Such BPI-  
derived peptides having at least one of the activities of BPI may be useful as  
25 antimicrobial agents (including anti-bacterial and anti-fungal agents), as endotoxin  
binding and neutralizing agents, and as heparin binding and neutralizing agents  
including agents for neutralizing the anticoagulant effects of administered heparin,  
for treatment of chronic inflammatory disease states, and for inhibition of normal or  
pathological angiogenesis. "Cationic BPI peptide" refers to a BPI peptide with a pI >  
30 7.0.

As used herein a "transformed bacterial host cell refers to a bacterial cell that  
contains recombinant genetic material or a bacterial cell that contains genetic

material required for expression of a recombinant product. The genetic material may be introduced by any method known in the art including transformation, transduction, electroporation and infection.

As used herein, a "vector" or "vector construct" refers to plasmid DNA that  
5 contains recombinant genetic material which may encode a recombinant product(s) and may be capable of autonomous replication in bacteria.

"Carrier protein" as used herein refers to a protein that can be expressed in bacteria and used as a fusion partner to a linked peptide or protein. Preferred carrier proteins are those that can be expressed at high yield and when used as a fusion  
10 partner can confer high level-expression to a linked peptide or protein. Particularly preferred carrier proteins are those that are expressed intracellularly as soluble or insoluble proteins, such as the D subunit of a human osteogenic protein ("Bone D"). Any known carrier protein may be utilized as a protein fusion partner, including, for example, ubiquitin, [see e.g., Pilon et al., *Biotechnol. Prog.* 13, 374-379 (1997)];  
15 staphylococcal protein A, [see e.g., Uhlén et al., *Gene* 23, 369:378 (1983) and Piers et al., *Gene* 134, 7-13 (1993)]; thioredoxin, [see e.g., LaVallie et al., *Bio/Technology* 11, 187-193 (1993)]; maltose binding protein, [see e.g., Tsao et al., *Gene* 169, 59-64 (1996)]; glutathione-s-transferase, [see e.g., Ray et al., *Bio/Technology* 11 64-70 (1993) and Piers et al., *Gene* 134, 7-13 (1993)]; prochymosin, [see e.g., Haught et al., *Biotechnology and Bioengineering* 57, 55-61 (1998)];  $\beta$ -galactosidase, [see e.g.,  
20 Kempe et al., *Gene* 39, 239-245 (1985)]; and gp 55 from T4, [see e.g., Gram et al., *Bio/Technology* 12, 1017-1023 (1994)]. A "cationic carrier protein" as used herein refers to a carrier protein having a pI (as calculated based on amino acid sequence or as measured in solution) greater than 7.0 and preferably greater than 8.0. Such  
25 proteins include (1) Bone D (pI 8.18) (SEQ ID NOS: 1 and 2) and (2) gelonin (pI 9.58) (see, e.g., U.S. Patent Nos. 5,416,202 and 5,851,802, hereby incorporated by reference in their entirety).

"Amino acid cleavage site" as used herein refers to an amino acid or amino acids that serve as a recognition site for a chemical or enzymatic reaction such that  
30 the peptide chain is cleaved at that site by the chemical agent or enzyme. Amino acid cleavage sites include those at aspartic acid - proline (Asp-Pro), methionine (Met), tryptophan (Trp) or glutamic acid (Glu). "Acid-sensitive amino acid cleavage

site" as used herein refers to an amino acid or amino acids that serve as a recognition site such that the peptide chain is cleaved at that site by acid. Particularly preferred is the Asp-Pro cleavage site which may be cleaved between Asp and Pro by acid hydrolysis.

5           Peptides derived from or based on BPI (BPI-derived peptides), are described in co-owned U.S. Patent No. 5,858,974 [WO 97/04008 (PCT/US96/03845)]; U.S. Patent Application Serial Nos. 08/504,841 and 09/119,858 [WO 96/08509 (PCT/US95/09262)]; U.S. Patent Nos. 5,652,332 and 5,856,438 [WO 95/19372 (PCT/US94/10427)]; U.S. Patent Nos. 5,733,872 and  
10 5,763,567 [WO 94/20532 (PCT/US94/02465)]; U.S. Patent Nos. 4,348,942; 5,639,727; 5,807,818; 5,837,678; and 5,854,214 [WO 94/20128 (PCT/US94/02401)]; the disclosures of all of which are incorporated herein by reference.

Other aspects and advantages of the present invention will be understood upon consideration of the following illustrative examples wherein Example 1  
15 addresses construction of fusion protein expression vector constructs; Example 2 addresses expression of recombinant fusion proteins; Example 3 addresses acid hydrolysis of bacterial cells or bacterial cell cultures and release of recombinant peptide; Example 4 addresses acid hydrolysis of bacterial cell cultures in fermentation vessels; Example 5 addresses acid hydrolysis of bacterial cells after  
20 removal of cell culture medium; Example 6 addresses recovery and purification of recombinant peptides from acid hydrolyzed bacterial cells; and Example 7 addresses biological activity assays of recombinant peptides.

### **EXAMPLE 1**

#### **25           Construction of Fusion Protein Expression Vectors**

##### **1.       Bacterial Expression Vector Construct pING4702**

A bacterial expression vector which would encode a peptide fusion protein, was constructed. This vector contains a sequence for a gene encoding  
30 subunit D of a human osteogenic protein ("Bone D") (*see*, amino acids 23 through 161 of SEQ ID NOS: 1 and 2), linked to a sequence encoding a linking sequence that includes the dipeptide Asp-Pro and a sequence encoding a peptide derived

from the sequence of BPI (SEQ ID NO: 3). This vector construct, pING4702, was prepared in several steps as described below.

First, two synthetic oligonucleotides were synthesized that encode a BPI-derived peptide, an Asp-Pro dipeptide and appropriate restriction enzyme

5 recognition sites for cloning. The oligonucleotides encoding this sequence were:

5'-GATCCACCGAAAGTGGGTTGGCTGATCCAGCTGTTCCACAAAA  
AGTAAAGC-3' (SEQ ID NO: 4)

5'-TCGAGCTTTACTTTTTGTGGAACAGCTGGATCAGCCAACCCACTTT  
CGGTG-3' (SEQ ID NO: 5)

10 Sixteen µg of each oligonucleotide were annealed in a 50 µL reaction in 100 mM NaCl, 10 mM Tris, pH 7.8, 1 mM EDTA for 10 minutes at 68°C, 30 minutes at 57°C, and followed by slow cooling to room temperature. The resulting annealed oligonucleotide fragment encodes an Asp-Pro -Pro sequence followed by sequence encoding a peptide with the 12 amino acid sequence of  
15 XMP.391, as described in U.S. Patent No. 5,851,802:

Asp Pro Pro Lys Val Gly Trp Leu Ile Gln Leu Phe His Lys Lys  
(SEQ ID NO: 6)

The annealed oligonucleotide fragment also contains restriction enzyme sites for cleavage by *Bam*HI at the 5' end and *Xho*I at the 3' end of sequence. The  
20 resulting annealed oligonucleotide was purified by centrifugation on a Chroma Spin 10 column (Clontech, Palo Alto, CA).

Second, DNA fragments from two plasmid vectors were prepared. Plasmid pIC100, a derivative of pBR322 and which includes the leader sequence of the *E. carotovora pelB* gene, described in U.S. Patent No. 5,416,202 (*see, e.g.*,  
25 Example 10) incorporated by reference, was digested with *Eco*RI and *Xho*I, and the large vector fragment of approximately 2836 bp, was purified. Plasmid pING3353, described in U.S. Patent No. 5,851,802, incorporated by reference, was digested with *Eco*RI and *Bam*HI and the approximately 550 bp fragment which encodes the *pelB*:Bone D protein was purified.

30 Third, the annealed oligonucleotide, the *Eco*RI to *Xho*I fragment from pING100 and the *Eco*RI to *Bam*HI fragment from pING3353 were ligated in 20 µL 50 mM Tris, pH 7.6, 10 mM MgCl<sub>2</sub>, 1 mM ATP, 1 mM DTT, 5% PEG-8000 with 3 Units T4 DNA Ligase for 16 hours at 4°C to generate the intermediate vector pING4700. Plasmid pING4700 confers ampicillin resistance and encodes  
35 the fusion protein Bone D-Asp-Pro-peptide.

Plasmid pING4700 was digested with *EcoRI* and *XhoI*, and the 604 bp fragment encoding the fusion protein was ligated to the approximately 5500 bp vector fragment from pING3217, as described in U.S. Patent 5,851,802, (see Example 1), that had been digested with *EcoRI* and *XhoI* in  $\mu\text{L}$  50 mM Tris, pH 7.6, 10 mM  $\text{MgCl}_2$ , 1 mM ATP, 1 mM DTT, 5% PEG-8000 with 3 Units T4 DNA Ligase for 16 hours at  $4\text{C}^\circ$ . The resulting plasmid, pING4702, encodes the Bone D-Asp-Pro-Pro-peptide fusion protein (SEQ ID NOS: 7 and 8) under the transcriptional control of the *araB* promoter. Plasmid pING4702 confers resistance to the antibiotic tetracycline.

10

## 2. Bacterial Expression Vector Construct pING4703

A second bacterial expression vector was constructed which encodes a peptide fusion protein containing Bone D (see, amino acid 23 through 161 of SEQ ID NOS: 1 and 2), the dipeptide Asp-Pro and a 25 amino acid peptide derived from the sequence of BPI (SEQ ID NO: 9). This vector construct, pING4703, was prepared as described below.

First, two synthetic oligonucleotides were synthesized that encode a BPI-derived peptide, an Asp-Pro-Pro sequence and appropriate restriction enzyme recognition sites for cloning. The oligonucleotides encoding this sequence were:

20 5'- CATTGGATCCACCGAAATGGAAGGCCAGTTTCGCTTTCTTAA  
GAAA TCGAAAGTGGGTTG -3' (SEQ ID NO: 10)  
5'- GGCTCTCGAGCTCTACTTTTATGAAACAGCAGGATCAGCCAACC  
CACTTTGATTCTTA -3' (SEQ ID NO: 11)

Sixteen  $\mu\text{g}$  of each oligonucleotide were annealed in a 50  $\mu\text{L}$  reaction in 25 100 mM NaCl, 10 mM Tris, pH 7.8, 1 mM EDTA for 10 minutes at  $68\text{C}^\circ$ , 30 minutes at  $57\text{C}^\circ$ , followed by slow cooling to room temperature. An aliquot of the annealed oligonucleotides was diluted into 10 mM Tris, pH 8.3, 50 mM KCl (300  $\mu\text{L}$  total volume) and filled-in with a reaction containing AmpliTaq (Perkin Elmer, Norwalk, CT), dATP, dGTP, dCTP and dTTP at  $72\text{C}^\circ$ . The resulting double-stranded fragment encoded the restriction sites *BamHI* and *XhoI* at the 5' and 3' ends, respectively, and encoded an Asp-Pro-Pro sequence followed by a sequence encoding a peptide with the 24 amino sequence of XMP.102, as described in U.S. Patent No. 5,851,802:

35 Asp Pro Pro Lys Trp Lys Ala Gln Phe Arg Phe Leu Lys Lys Ser Lys Val Gly Trp  
Leu Ile Leu Leu Phe His Lys Lys (SEQ ID NO: 12)

The double-stranded fragment was digested with *Bam*HI and *Xho*I, and ligated to both the approximately 5500 bp *Eco*RI to *Xho*I vector fragment from pING3217, and the approximately 550 bp *Eco*RI to *Bam*HI fragment of pING3353 in 20  $\mu$ L 50 mM Tris, pH 7.6, 10 mM MgCl<sub>2</sub>, 1 mM ATP, 1 mM dithiothreitol, 5% PEG-8000 with 1 Unit T4 DNA Ligase for 16 hours at 4C°. The resulting plasmid, pING4703, encodes the Bone D-Asp-Pro-Pro-peptide fusion protein (SEQ ID NOS: 13 and 14) under the transcriptional control of the *araB* promoter. Plasmid pING4703 confers resistance to the antibiotic tetracycline.

10

## **EXAMPLE 2**

### **Expression of Recombinant Fusion Proteins**

Expression of a recombinant product under control of the *araB* promoter was evaluated as follows. Expression vector constructs were transformed into *E. coli* E104 (deposited as ATCC 69009; ATCC 69008; ATCC 69101; ATCC 69102; ATCC 69103; ATCC 69104; ATCC 69331; ATCC 69332; ATCC 69333, each containing a gelonin-encoding plasmid) and tetracycline resistant colonies were selected. Bacterial cultures from these colonies were grown at 37°C in TYE medium (15 g Tryptone, 10 g Yeast Extract, 5 g NaCl per liter) supplemented with 15  $\mu$ g/mL of tetracycline. For storage of bacterial cells prior to growth in a fermentor, bacterial cultures (1 to 2 mL) were frozen in TYE medium supplemented with 15% glycerol and stored at -20°C. To initiate production of recombinant product, a vial of cells containing the product expression vector was thawed, and inoculated into 100 mL of GMM culture medium as described below and grown to approximately 200 Klett Units, then inoculated into either a 14 L or 35 L fermentor. Each fermentor contained a minimal salts medium with glycerol as a carbon source (Glycerol Minimal Medium, GMM). The 14 L or 35 L fermentor vessel initially contained approximately 7 L or 20 L, respectively, of GMM which contains the following ingredients per liter:

	<u>Autoclaved Ingredients</u>	
	(NH <sub>4</sub> ) <sub>2</sub> SO <sub>4</sub>	12 g
	KH <sub>2</sub> PO <sub>4</sub>	1.57 g
5	K <sub>2</sub> HPO <sub>4</sub>	14.1 g
	MgSO <sub>4</sub> ·7H <sub>2</sub> O	0.28 g
	H <sub>3</sub> PO <sub>4</sub> (Conc.)	3 mL
	Antifoam	1 mL
	Biotin	0.0012 g
10	Yeast Extract	4.6 g
	Glycerol	18.5 g
	<u>Filter sterilized ingredients</u>	
15	CaCl <sub>2</sub> ·2H <sub>2</sub> O (10% w/v)	1 mL
	Trace D Solution*	16 mL
	Thiamine HCl (10% w/v)	0.1 mL
	Nicotinic Acid (1% w/v)	2 mL
20	*Trace D solution is composed of:	
	FeCl <sub>3</sub> ·6H <sub>2</sub> O	6.480 g
	ZnSO <sub>4</sub> ·7H <sub>2</sub> O	1.680 g
	MnCl <sub>2</sub> ·4H <sub>2</sub> O	1.200 g
25	Na <sub>2</sub> MoO <sub>4</sub> ·2H <sub>2</sub> O	0.576 g
	CuSO <sub>4</sub> ·5H <sub>2</sub> O	0.240 g
	CoCl <sub>2</sub> ·6H <sub>2</sub> O	0.240 g
	H <sub>3</sub> BO <sub>3</sub>	0.720 g
	H <sub>3</sub> PO <sub>4</sub> (Conc.)	96.0 mL
30	H <sub>2</sub> O (Batch Volume)	2.0 L

The fermentor was then inoculated with the bacterial seed culture, and was maintained at pH 6.0 and 32°C with 10 L/min. air and agitation at 1000 rpm. When nutrients became limiting (as judged by an increase in the dissolved oxygen, DO, to approximately 100%), the culture was fed with additional nutrients until the culture reached an optical density (OD<sub>600</sub>) of approximately



100. Culture feed rate was controlled to maintain the DO to a setpoint of 20%. Specifically, the culture was fed with the first feed:

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Autoclaved ingredients per liter of feed:

5	Glycerol	700 g
	MgSO <sub>4</sub> ·7H <sub>2</sub> O	10 g
	Biotin	0.01 g

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Filtered ingredients per liter of feed

10	CaCl <sub>2</sub> ·2H <sub>2</sub> O (10% w/v)	35 mL
	Thiamine HCl (10% w/v)	3.5 mL
	Nicotinic Acid (1% w/v)	7 mL

---

15           The culture was induced by gradient induction at an OD of approximately 100 with a second feed containing the inducing agent L-arabinose. Specifically, the second feed was:

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Autoclaved ingredients per liter of feed:

20	Glycerol	700 g
	MgSO <sub>4</sub> ·7H <sub>2</sub> O	10 g
	Biotin	0.01 g
	Arabinose	60 g/L

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Filtered ingredients per liter of feed

25	CaCl <sub>2</sub> ·2H <sub>2</sub> O (10% w/v)	35 mL
	Thiamine HCl (10% w/v)	3.5 mL
	Nicotinic Acid (1% w/v)	7 mL

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30

The cultures were harvested 23-26 hours post induction.

The cells may be separated from the culture medium with a 0.2 µm hollow fiber cartridge, 10 ft. <sup>2</sup> (Microgon, Laguna Hills, CA) as described in Examples 3 and 5 below. Alternatively, the fermentation broth (*i.e.*, culture medium with cells) may be used directly in the fermentation vessel or removed from the fermentor for acidification and further processing as described in Examples 4 and

5 below. Example 6 below describes the recovery and purification of recombinant peptides.

### EXAMPLE 3

#### 5 Acid Hydrolysis of Bacterial Cells and Release of Recombinant Peptide

##### 1. Peptide release from bacterial cells

Previously, inclusion bodies were isolated. Acid treatment of the isolated inclusion bodies resulted in the hydrolysis of the aspartyl-prolyl bond (Asp-Pro) between the Bone D protein and a recombinant peptide by incubation in dilute acid at elevated temperatures (*see, e.g.* Example 3 of U.S. Patent No. 5,851,802). In these experiments, bacterial cells were directly acidified in an attempt to lyse the cells and hydrolyze the inclusion bodies directly to release the peptide. This was done by diluting cells in dilute acid at elevated temperature. *E. coli* E104 containing plasmid pING4702 was grown in a 10 L fermentor and induced with arabinose. After termination of the fermentor run, bacterial cells were separated from the majority of the culture supernatant with a 0.2  $\mu$ m hollow fiber cartridge (Microgon, 10ft<sup>2</sup>) and frozen. Cells obtained from the fermentor were thawed and incubated under acidic conditions for 4 hours at 85°C as follows in Table 1.

TABLE 1

Sample	Cell Paste	Incubation Condition
A	1 gram	10 mL of 30 mM HCl
B	1 gram	10 mL of 30 mM HCl, 5 mM EDTA
C	1 gram	10 mL of 30 mM HCl, 5 mM EDTA, 1% Triton X-100
D	1 gram	10 mL of 30 mM HCl, 5 mM EDTA, 8 M urea

As a control, approximately 1 gram of cells were lysed with lysozyme and inclusion bodies were isolated prior to acid hydrolysis according to prior methods (*see, e.g.*, Example 3 of U.S. Patent No. 5,851,802). These cells were suspended in 10 mL of 100 mM Tris, 5 mM EDTA, pH 8.0. The slurry was incubated on ice for 15 minutes, and 1 mL of 10 mg/mL lysozyme was added and incubated on ice for 20 minutes. To disrupt the lysozyme treated cells, the slurry was sonicated 4

times for 10 seconds each at the highest setting using a Sonic U sonicator (B. Braun Biotech Inc., Allentown, PA). The lysed cells were centrifuged at 13,000 rpm in a JA20 rotor for 25 minutes. The inclusion body pellet was then incubated in 10 mL of 30 mM HCl for 4 hours at 85°C.

5 Prior to incubation at 85°C, the pH of all samples was adjusted to pH 2.5 with HCl, except for the sample containing urea which was adjusted to pH 3.0. After incubation, the samples were centrifuged at 13,000 rpm in a JA20 rotor for 25 minutes to separate soluble from insoluble material. The amount of released peptide in the supernatant from each sample was evaluated by HPLC using a

10 Beckman Coulter (Fullerton, CA) instrument with a Shimadzu Scientific Instruments (Columbia, MD) auto injector and a Vydac (Hesperia, CA) C18 (#218TP54) column. Solvent A was 10% acetonitrile/0.1% TFA; solvent B was 90% acetonitrile/0.1% TFA. The column was run with an 20-40% B gradient over 20 minutes at a flow rate of 1 mL/minute with peptide detection at 229 nm.

15 The concentration of peptide in the supernatant was as follows in Table 2.

**TABLE 2**

	Sample	Concentration (mg/mL)	% of Control
	Purified Inclusion bodies (Control)	0.292	100
A	10 mL of 30 mM HCl	0.223	76.4
B	10 mL of 30 mM HCl, 5 mM EDTA	0.218	74.7
C	10 mL of 30 mM HCl, 5 mM EDTA, 1% Triton X-100	0.287	98.3
D	10 mL of 30 mM HCl, 5 mM EDTA, 8 M urea	0	0

20 These data demonstrate for the first time that peptide could be released directly from cells by incubation of the bacterial cells in dilute acid while the majority of other proteins remain insoluble.

## 2. Timecourse of peptide release from bacterial cells.

25 The results described above demonstrated that peptide was released from a Bone D-peptide fusion protein containing an acid sensitive Asp-Pro peptide linker by direct hydrolysis of cells in dilute acid. Studies were performed to examine the

timecourse for hydrolysis. A sample of the same concentrated, frozen cell sample described above was used for additional studies. Approximately 2 grams of cell paste was diluted with 20 mL of water and concentrated HCl was added to bring the pH to 2.5. The sample was incubated at 85°C, and samples were removed  
5 periodically for quantitation. Each sample was centrifuged to remove insoluble material, and the supernatant was assayed for released peptide by HPLC. The concentration of peptide in the soluble fraction was as follows in Table 3.

TABLE 3

10

Time (Hours)	Concentration by HPLC (mg/mL)
0	0
0.5	0.02
1	0.03
2	0.056
3	0.082
4	0.142
5	0.184
6	0.219
7	0.267

Thus, as shown in Table 3, the amount of peptide in the soluble fraction was still increasing at the end of the seven hour timecourse.

In additional studies, a cell sample of bacterial cells in cell culture media  
15 was incubated at pH 2.15 to evaluate the timecourse of peptide release from cells that had not been previously concentrated and frozen. Specifically, 40 mL of bacterial cells in fermentation broth (fermentation culture of *E. coli* E104 containing pING4702) at the end of the fermentor process as described in Example 2 was directly adjusted to pH 2.15 by adding 500 µL of concentrated  
20 HCl, to a final concentration of approximately 150 mM. The sample was incubated at 85°C and every hour a sample was removed, centrifuged, and the supernatant was evaluated for peptide by HPLC. The amount of peptide released over time was as follows in Table 4.

TABLE 4

Time (Hours)	Concentration by HPLC (mg/mL)
0	0
1	0.062
2	0.218
3	0.321
4	0.350
5	0.366
6	0.392
7	0.432
8	0.401
23	0.298

5           At pH 2.15 and using cells directly in the fermentation medium, maximum release of peptide occurred by seven hours at 85°C, after which the amount of released peptide decreased.

          Additional studies demonstrated that dilute H<sub>2</sub>SO<sub>4</sub> and HNO<sub>3</sub> could also release soluble peptide from bacterial cells and bacterial cells in cell culture media  
10   (*i.e.*, fermentation cultures). In studies with H<sub>2</sub>SO<sub>4</sub>, two 20 mL samples of bacterial cells in fermentation broth as described in Example 4 (fermentation culture of *E. coli* E104 containing pING4702) were collected after completion of a bacterial fermentation, and they were acidified to pH 2.4. One sample was adjusted to pH 2.4 with HCl and the other was adjusted to pH 2.4 with H<sub>2</sub>SO<sub>4</sub>.  
15   Each sample was incubated at 85°C, a sample was removed every hour for seven hours and the amount of soluble peptide in each sample was analyzed by HPLC. The concentration of peptide in each aliquot is shown in the following Table 5.

TABLE 5

Sample Time (Hour)	Peptide Concentration by HPLC (mg/mL)	
	HCl Hydrolysis	H <sub>2</sub> SO <sub>4</sub> Hydrolysis
0	0	0
1	0.034	0.036
2	0.130	0.116
3	0.185	0.162
4	0.231	0.209
5	0.281	0.248
6	0.295	0.275
7	0.303	0.273

- 5 In studies with HNO<sub>3</sub>, a sample of cells in bacterial fermentation broth as described in Example 4 was incubated with nitric acid. Specifically, 20 mL of cells were adjusted to pH 2.2 with nitric acid and incubated at 85°C. Samples were removed periodically and the concentration of recombinant peptide in the soluble fraction was determined by HPLC. The concentration of peptide in each
- 10 aliquot is shown in the following Table 6.

TABLE 6

Sample Time (Hour)	Peptide Concentration by HPLC (mg/mL) after Hydrolysis with HNO <sub>3</sub>
0	0
1	0.093
2	0.146
4.5	0.308
6	0.325

- 15 These additional studies demonstrate that acids such as nitric acid, that are less corrosive to stainless steel materials used in fermentation vessels, are useful in the improved methods of the invention.

**EXAMPLE 4****Acid Hydrolysis of Bacteria Directly in a Fermentation Vessel**

Since peptide was released from bacterial cells and cell cultures by direct  
5 incubation of cells in acid as described in Example 3, studies were done to  
evaluate if soluble peptide could be recovered directly from a bioreactor at the end  
of the fermentation process when contents of the fermentor were acidified and  
heated in place. In initial studies, *E. coli* E104 containing pING4702 was grown  
in a 35 L fermentor as described in Example 2. The first feed solution was  
10 introduced in the fermentor at 20.5 hours after inoculation, and the culture was  
induced with the second feed when the OD600 had reached 97.2. At 62.5 hours  
after induction, 10% HCl was added to the fermentor in 50 mL aliquots until the  
pH of the fermentor had reached approximately 2.28. In total, 990 mL of acid was  
added to the approximately 26 L of fermentation product in the fermentor. After  
15 reducing the pH, the temperature setpoint on the fermentor was increased to 85°C,  
and samples were removed from the fermentor periodically thereafter for six  
hours. The contents of the vessel were mixed during the reaction with the  
fermentor impellers. HPLC analysis of the soluble material in the samples  
revealed that the concentration of the peptide leveled off between four and five  
20 hours. The concentration of peptide was as shown in the following Table 7.

**TABLE 7**

<b>Sample Timepoint (Hours)</b>	<b>Peptide Concentration by HPLC (mg/mL)</b>
0	0
1	0.185
2	0.294
3	0.334
4	0.353
5	0.353
6	0.370

25 In additional studies, *E. coli* E104 containing pING4702 was grown in a  
35 L fermentor to an OD600 of 89, induced with the second feed containing  
arabinose and grown for 24 hours. A 10% HCl solution was added to bring the

culture pH to approximately 2.3, and the temperature was raised to 85°C for 5.5 hours. The concentration of peptide in the soluble fraction was 0.332 mg/mL.

5

**EXAMPLE 5****Acid Hydrolysis of Bacteria After Removal of the Cell Culture Medium**

*E. coli* E104 containing pING4703 was grown in a 14 L fermentor as described in Example 2, and 10 mL of the fermentation culture was adjusted to pH 2.2 with concentrated HCl. The sample was incubated at 85°C, and samples were taken every few hours and analyzed for peptide in the supernatant by HPLC, using the same method as described in Example 3 for quantitation of peptide from the product encoded by pING4702. The results from this study are shown in the following Table 8.

15

**TABLE 8**

Time at 85°C	Peptide Concentration mg/mL
4	0.018
6	0.011
7	0.004

These peptide titers were much lower than what was obtained with cultures of *E. coli* E104 (pING4702) as described in Examples 3 and 4, and lower than the titer obtained when *E. coli* E104 (pING4703) was lysed by sonication after incubation with lysozyme by the process described in Example 3. *E. coli* E104 (pING4703) lysed by sonication after lysozyme treatment had a titer of approximately 0.46 mg/mL in the soluble fraction.

25

In additional studies, samples of the bacterial cell culture both before and after acid hydrolysis at 85°C were analyzed by SDS-PAGE. The results demonstrated that the fusion protein of Bone D and peptide had been hydrolyzed by acid. An experiment was executed to determine if the cell culture medium in the hydrolysis reaction had an impact on the ability to recover recombinant peptide in the soluble fraction, since a prominent band at the position of Bone D was apparent in the hydrolyzed sample, while very little intact fusion protein was detected. Cell paste from the fermentation of *E. coli* E104 (pING4703) was

30



prepared by centrifugation, and 1 g of cell paste was suspended in: 7 mL H<sub>2</sub>O; 7 mL of 5 mM EDTA; or 7 mL of cell-free fermentation broth from the same bacterial fermentor. Each sample was adjusted to pH 2.2 with concentrated HCl, and incubated at 85°C. The amount of recombinant peptide in the soluble fraction was measured over time by HPLC. The results are shown in the following Table 9.

TABLE 9

Time (Hours) at 85°C	H <sub>2</sub> O Sample	5 mM EDTA Sample	Medium Sample
Peptide Concentration by HPLC (mg/mL)			
1	0.052	0.043	0
2	0.184	0.187	0.011
4	0.269	0.284	0.009
6	0.267	not determined	0.003
8	0.255	not determined	not determined

These data demonstrated that the recombinant peptide was soluble when the cells were hydrolyzed in water or 5 mM EDTA, but did not become soluble in the fermentation medium after acid hydrolysis.

Further studies were performed to determine if recombinant peptide was insoluble in acid after hydrolysis from Bone D, and could be released from the insoluble material in detergents or chaotropic salts. Three 1 gram samples of cell paste from *E. coli* E104 (pING4703) were suspended in 7 mL of 100 mM Tris, 5 mM EDTA, pH 8.0, and one 1 gram sample of cell paste was suspended in 7 mL of cell-free culture medium from the *E. coli* fermentation. To one of the samples suspended in Tris buffer, 1 mL of 10 mg/mL lysozyme was added, the sample was incubated on ice and sonicated as described in Example 3. The pH of all four samples was adjusted to approximately pH 2.0 with concentrated HCl, and the samples were incubated at 85°C for 4 hours. By HPLC, the amount of peptide released into the soluble fraction from the four samples was as follows in Table 10.

TABLE 10

Sample	Suspension buffer	Peptide Concentration mg/mL
1	100 mM Tris, 5 mM EDTA	0.594
2	100 mM Tris, 5 mM EDTA	0.618
3	Medium	0
4	100 mM Tris, 5 mM EDTA + Lysozyme and sonication	0.519

- 5 Thus, peptide did not appear in the soluble fraction in Sample 3 after acid hydrolysis. To determine if peptide could be released from the insoluble material, the pellet from Sample 3 was washed sequentially with 7 mL of buffer containing Triton X-100, urea, guanidine hydrochloride or SDS. The amount of peptide released from the pellet was as follows in Table 11.

10

TABLE 11

Wash Buffer	Peptide concentration (mg/mL) in the Wash Buffer	Total Peptide Released mg peptide/gram of cells
1% Triton X-100 in 10 mM sodium phosphate, pH 7.0	0.01	0.08
3% Triton S-100 in 10 mM sodium citrate, pH 3.0	0	0
4 M urea in 10 mM sodium citrate, pH 3.0	0.04	0.29
8 M urea in 10 mM sodium citrate, pH 3.0 - first wash	0.08	0.55
8 M urea in 10 mM sodium citrate, pH 3.0 - second wash	0.07	0.49

Wash Buffer	Peptide concentration (mg/mL) in the Wash Buffer	Total Peptide Released mg peptide/gram of cells
8 M urea in 10 mM sodium citrate, pH 3.0 - third wash for 15 hours	0.06	0.39
6 M guanidine hydrochloride	0.12	0.87
4% SDS	0	0
		Total in all washes: 2.67

These results demonstrated that the peptide could be recovered from the insoluble material by washing in buffers containing urea or guanidine hydrochloride. The peptide was therefore not degraded by the hydrolysis condition, but is rendered insoluble by media components. The total amount of material recoverable in all washes was 2.67 mg per gram of cells, compared to 4.16 mg/g and 3.63 mg/g recovered directly from the soluble material in Samples 1 and 4, respectively. Thus, for some bacterial cell cultures, the bacterial cells may be preferentially removed from the media and the bacterial cells may be acidified according to Example 3. For other bacterial cell cultures, the fermentation broth (bacterial cells in cell culture/fermentation media) may be directly acidified according to Examples 3 and 4.

#### EXAMPLE 6

##### Recovery and Purification of Recombinant Peptide from Acid Hydrolyzed Cells

##### 1. Recovery

The invention provides methods for the recovery of peptides in the soluble fraction after acid hydrolysis of cells while the large majority of other bacterial proteins, the carrier protein, and other impurities remain in the insoluble fraction. The soluble and insoluble material can be separated by centrifugation, filtration or any other suitable separation method. Any variety of centrifuge can be used to separate these materials and a variety of filtration devices, systems and methods can also be used. A variety of such filtration devices, systems and methods were used to separate soluble and insoluble materials including dead end (depth)

filtration and tangential flow filtration. A summary of the results of exemplary filtration studies to separate soluble and insoluble material by filtration is presented in the following Table 12.

5

TABLE 12

Filtration Method	Sample Analyzed	Filter Description	Through-put	Permeate description	Recovery
Depth	Previously frozen cell paste suspended in water and acid hydrolyzed	Seitz 900 SD/SDC, 1 ft <sup>2</sup> , 8 $\mu$ m nominal retention	1.5 L	Clear	72%
Depth preceded by 1 $\mu$ m bag filter	Previously frozen cell paste suspended in 4 volumes of water and acid hydrolyzed	Seitz SD250, 1 ft <sup>2</sup> , 4 $\mu$ m nominal retention	5.5 L	Clear	94%
Depth	Acid hydrolysate prepared directly in a 35 L fermentor	Cuno Zeta Plus 01A, 1 ft <sup>2</sup> , 7 $\mu$ m nominal retention	1.7 L	Cloudy	ND
Depth preceded by 1 $\mu$ m bag filter	Acid hydrolysate prepared directly in a 35 L fermentor	Cuno 30 SP, 1 ft <sup>2</sup> , 0.6 $\mu$ m nominal retention	1.2 L	Cloudy	ND
Depth with and without Celite filter aid	Previously frozen cell paste suspended in 3.8 volumes of water and acid hydrolyzed	Cuno Zeta Plus 01A, 28 cm <sup>2</sup>	27 mL with no Celite; > 50 mL with Celite (HP <sup>3</sup> 1000)	Slightly cloudy	ND

Filtration Method	Sample Analyzed	Filter Description	Through-put	Permeate description	Rec very
Celite filter aid in horizontal pressure leaf vessel	Acid hydrolysate prepared directly in a 35 L fermentor	Celite (Hy-flo) precoat, 600 cm <sup>2</sup>	2 L/ min	Clear	73%
Tangential Flow	Previously frozen cell paste suspended in 3.8 volumes of water and acid hydrolyzed	Sartorius 0.2 cutoff filter, 0.1 m <sup>2</sup>	ND	Cloudy	ND
Tangential Flow	Previously frozen cell paste suspended in 3.8 volumes of water and acid hydrolyzed	300 kDa MWCO, 0.1 m <sup>2</sup>	150 mL/min	Clear	81%

ND - not determined

Seitz filters are products of SWK Filtration Incorporated, Petaluma, CA.

Cuno filters are products of Cuno, Meriden, CT.

Celite is a product of World Minerals, Lompoc, CA.

5 Sartorius filters are products of Sartorius, Edgewood, NY.

These results demonstrate that a variety of filtration devices, systems and methods can be successfully employed to separate the soluble and insoluble material.

10

## 2. Purification

Following fermentation of *E. coli* E104 containing pING4702 as described in Example 2, bacterial cells in the unprocessed fermentation broth were hydrolyzed in dilute HCl. Specifically, 40 mL of fermentation broth was adjusted to pH 2.15 with concentrated HCl. The sample was incubated at 85°C for 5.5 hours. The hydrolyzed cells were then centrifuged to remove insoluble material, and the supernatant was adjusted to pH 3.0 by adding 500 mM sodium citrate dropwise.

15

An SP Sepharose (Amersham-Pharmacia, Piscataway, NJ) column, 2.5 x 4.4 cm containing 21.6 mL, was equilibrated in 10 mM sodium citrate, pH 3.0 and

20

the sample was loaded. The column was washed with 10 mM sodium citrate , pH 3.0 buffer and then 10 mM sodium phosphate, pH 7.0 until the pH of the column effluent reached 7. The column was then washed in 10 mM sodium phosphate, 150 mM NaCl pH 7.0. The column was eluted in 10 mM sodium phosphate, 800 mM NaCl pH 7.0 and then the column was stripped with 10 mM sodium phosphate, 2 M NaCl. The SP Sepharose eluate was diluted with one volume of 10 mM sodium phosphate, 3 M ammonium sulfate, pH 7.0.

A Butyl Sepharose (Amersham-Pharmacia) column, 1 x 4 cm containing 3.1 mL, was equilibrated with 10 mM sodium phosphate, 1.5 M ammonium sulfate, pH 7.0, and the sample was loaded. The Butyl Sepharose column was washed with 10 mM sodium phosphate, 1.1 M ammonium sulfate, pH 7.0, and then eluted with 10 mM sodium phosphate, 0.4 M ammonium sulfate, pH 7.0. The column was striped with 10 mM sodium phosphate, pH 7.0

The peptide concentration in each of the fractions from the SP Sepharose and Butyl Sepharose columns was followed by HPLC analysis. The sample volumes, peptide concentrations and percent recovery was as follows in Table 13.

TABLE 13

Sample	Volume (mL)	Concentration (mg/mL)	Total mg	% Yield
SP Sepharose load	24	0.399	9.58	100
First SP Sepharose wash	75	0	0	0
Second SP Sepharose wash	33	0	0	0
SP Sepharose eluate	50	0.165	8.25	86.1
SP Sepharose strip	13	0.036	0.47	4.9
Butyl Sepharose load	94	ND	ND	ND
Butyl Sepharose flow through	95	0	0	0
Butyl Sepharose wash	13	0.008	0.1	1
Butyl Sepharose eluate	28	0.262	7.34	76.6
Butyl Sepharose strip	4	0.014	0.06	0.6

ND -Not Determined

A Superdex 30 (Amersham-Pharmacia) column, 1.6 x 53 cm containing 107 mL, was equilibrated in 5 mM sodium acetate, 150 mM NaCl, pH 5.0. Eight

mL of Butyl Sepharose eluate was loaded onto the Superdex 30 gel filtration column, and the column was run with 5 mM sodium acetate, 150 mM NaCl, pH 5.0. After 32 mL had flowed through the column, 3 mL fractions were collected. Fractions 12-19 were pooled and had a volume of approximately 20 mL. The  
5 concentration of recombinant peptide in the Superdex 30 pool was 0.107 mg/mL for a recovery of 102% from the previous step, and the overall recovery from the acid hydrolysate of cells was 76.6%. The final peptide purity was 97.4%.

#### EXAMPLE 7

##### 10 Biological Activity Assays of Recombinant Peptides

A variety of recombinant peptides, including those BPI-derived peptides comprising the sequences listed in U.S. Patent No. 5,851,802 incorporated by reference, may be produced by recombinant methods of the invention and tested  
15 for biological activity by known activity assays. Assays for antimicrobial activity (both anti-fungal and anti-bacterial activity) may be performed, including radial diffusion assays. Assays, with a variety of fungal and bacterial cells, including those described in U.S. Patent No. 5,851,802 (see Example 6), may be conducted using recombinant peptides produced according to the invention.

20 For example, studies were performed to evaluate the antifungal activity of the recombinant peptide from pING4702 purified according to Example 6 in a broth microdilution assay using four strains of *C. albicans*, *C. glabrata* and *S. cerevisiae*. A similar peptide, XMP.391, that was chemically synthesized, was included in the assay as a positive control. To perform the broth microdilution  
25 assay, the fungal cultures were grown overnight at 30°C in YPD medium (1% yeast extract, 2% peptone, and 2% dextrose). A 400 fold dilution of each culture in YPD was then made, and grown at 30°C for 8 hours. Three mL of each culture were collected by centrifugation and suspended in 0.9% NaCl to an A600 of about 0.3. These cultures were further diluted to  $1 \times 10^4$  CFU/mL in Sabouraud  
30 dextrose broth (6 mL). Recombinant peptide was in 5 mM acetate, 150 mM NaCl, pH 5.0 at a concentration of about 2 mg/mL. Synthetic peptide was at about 1 mg/mL. Samples were serially diluted and added to microtiter plates

containing the cultures. Plates were incubated at 30°C for 48 hours before growth inhibition was measured. Results from this assay were as follows in Table 14.

TABLE 14

5

<u>Strain</u>	<u>Synthetic Peptide XMP.391</u> (Concentration that gives 95% inhibition ( $\mu$ M))	<u>Recombinant Peptide</u> (Concentration that gives 95% inhibition ( $\mu$ M))
<i>C. albicans</i> SLU1	13.5	16
<i>C. albicans</i> 10231	16	30
<i>C. albicans</i> 14053	16	16
<i>C. albicans</i> 26555	16	30
<i>C. glabrata</i> 2001	30	30
<i>S. cerevisiae</i> 9763	2.0	7.5

Additionally or alternatively, assays may be performed to assess the endotoxin binding and neutralizing activity of the recombinantly produced peptides, by a variety of known assays, including those described in co-owned  
 10 U.S. Patent Nos. 5,733,872 and 5,763,567 [WO 94/20532 (PCT/US94/02465)];  
 5,652,332 and 5,856,438 [WO 95/19372 (PCT/US94/10427)]; 5,858,974 [WO  
 96/08509 (PCT/US95/09262) and WO 97/04008 (PCT/US96/03845)];  
 incorporated by reference in their entirety.

Additionally or alternatively, assays may be performed to assess the  
 15 heparin binding and neutralizing activity of the recombinantly produced peptides  
 by a variety of known assays, including assays as described in U.S. Patent Nos.  
 5,348,942; 5,639,727; 5,807,818; 5,837,678; and 5,854,214 [WO 94/20128  
 (PCT/US94/02401)]; 5,733,872 and 5,763,567 [WO 94/20532 (PCT/US94/  
 02465)]; 5,652,332 and 5,856,438 [WO 95/19372 (PCT/US94/10427)];  
 20 incorporated by reference in their entirety.

It should be understood that the foregoing disclosure emphasizes certain specific embodiments of the invention and all modifications or alternatives equivalent thereto are within the spirit and scope of the invention as set forth in



the appended claims. In particular, numerous modifications and variations in the practice of the invention are expected to occur to those skilled in the art upon consideration of the foregoing description on the presently preferred embodiments thereof. Consequently, the only limitations which should be placed upon the

5    scope of the present invention are those that appear in the appended claims.

## WHAT IS CLAIMED IS:

1. In a method for obtaining a peptide from bacterial cells after expression inside the cells of a fusion protein, wherein the fusion protein  
5 comprises the peptide, a carrier protein and an acid-cleavable site between the peptide and the carrier protein, the improvement comprising: treating the bacterial cells with acid under conditions sufficient in a single step to disrupt or lyse the cells and release the peptide from the fusion protein.
- 10 2. The method of claim 1 with the additional step of obtaining the released peptide separated from the disrupted or lysed cells.
3. The method of claim 2 wherein the released peptide is separated from the disrupted or lysed cells by a separation device.
- 15 4. The method of claim 3 wherein the separation device is a centrifugation device.
5. The method of claim 3 wherein the separation device is a filtration  
20 device.
6. The method of claim 1 wherein the acid-cleavable site in the fusion protein is Asp-Pro.
- 25 7. The method of claim 1 wherein the carrier protein is expressed as an insoluble protein inside the bacterial cells.
8. The method of claim 7 wherein the carrier protein is the D subunit of human osteogenic protein.
- 30 9. The method of claim 1 wherein the bacterial cells are in cell culture media for the acid treatment.

10. The method of claim 1 wherein the bacterial cells have been separated from cell culture media for the acid treatment.

5 11. The method of claim 1 wherein the bacterial cells are in cell culture media in a fermentation vessel for the acid treatment.

12. In a method for obtaining a peptide from bacterial cells after expression inside the cells of a fusion protein, wherein the fusion protein  
10 comprises the peptide, a carrier protein and an acid-cleavable site between the peptide and the carrier protein, the improvement comprising:  
(a) treating the bacterial cells with acid under conditions sufficient to disrupt or lyse the cells and release the peptide from the fusion protein,  
(b) separating soluble material from insoluble material after step (a),  
15 and  
(c) recovering the released peptide in the soluble material after step (b).

13. The method of claim 12 wherein the soluble material is separated  
20 from the insoluble material by a separation device.

14. The method of claim 13 wherein the separation device is a centrifugation device.

25 15. The method of claim 13 wherein the separation device is a filtration device.

16. The method of claim 12 wherein the acid-cleavable site in the fusion protein is Asp-Pro.

30

17. The method of claim 12 wherein the carrier protein is expressed as an insoluble protein inside the bacterial cells.

18. The method of claim 17 wherein the carrier protein is the D subunit of human osteogenic protein.

5 19. The method of claim 12 wherein the bacterial cells are in cell culture media for the acid treatment.

20. The method of claim 12 wherein the bacterial cells have been separated from cell culture media for the acid treatment.

10

21. The method of claim 12 wherein the bacterial cells are in cell culture media in a fermentation vessel for the acid treatment.

-1-

## SEQUENCE LISTING

<110> Better, Marc D.  
Gavit, Patrick D.

<120> Improved Methods for Recombinant Peptide Production

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 Ile Pro Asp Tyr Ser Asp Ser Phe Lys Ile Lys His Leu Gly Lys Gly  
 35 40 45



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 Ser Ala Asp Leu Lys Leu Gly Ser Asn Pro Thr Ser Gly Lys Pro Thr  
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# INTERNATIONAL SEARCH REPORT

International Application No.

PCT/US 00/07148

## A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 C12N15/12 C12N15/62 C07K14/47 C07K14/51 C12P21/06

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C12N C07K C12P

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, WPI Data, PAJ, MEDLINE, BIOSIS, EMBASE

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO 97 35009 A (BETTER MARC D ;XOMA CORP (US)) 25 September 1997 (1997-09-25) cited in the application the whole document	1,6-8, 12,16-18
A	GRAM H ET AL.: "A novel approach for high level production of a recombinant human parathyroid hormone fragment in Escherichia coli" BIO/TECHNOLOGY, vol. 12, no. 10, October 1994 (1994-10), pages 1017-1023, XP000922604 cited in the application abstract	1,6,7, 12,16,17

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

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- \*O\* document referring to an oral disclosure, use, exhibition or other means
- \*P\* document published prior to the international filing date but later than the priority date claimed

- \*T\* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- \*X\* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- \*Y\* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
- \*S\* document member of the same patent family

Date of the actual completion of the international search

21 August 2000

Date of mailing of the international search report

31/08/2000

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## INTERNATIONAL SEARCH REPORT

Int. Jonal Application No

PCT/US 00/07148

## C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	AUSUBEL F M ET AL.: "Chemical cleavage of fusion proteins by hydrolysis at low pH" CURRENT PROTOCOLS IN MOLECULAR BIOLOGY, 1994, page 16.4.13 XP002145208 the whole document	1,6,12, 16
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A	EP 0 213 472 A (SYNTEX INC) 11 March 1987 (1987-03-11) page 32, line 28 -page 34, line 32 examples 19,20 claims 36-39	1,6,12, 16
T	GAVIT P ET AL.: "Production of antifungal recombinant peptides in Escherichia coli" JOURNAL OF BIOTECHNOLOGY, vol. 79, no. 2, 28 April 2000 (2000-04-28), pages 127-136, XP000922726 the whole document	1-21

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Information on patent family members

International Application No

PCT/US 00/07148

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
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